

Editorial Commentary: Peripheral Blood Stem Cells Mobilization Using Granulocyte Colony-Stimulating Factor for Articular Cartilage Injuries: Wake Them Up and Make Them Come to You!



Jorge Chahla, M.D., Ph.D., Editorial Board, and Safa Gursoy, M.D., Ph.D.

Abstract: Articular cartilage injuries constitute a prevalent musculoskeletal problem in the general population. Restorative cartilage procedures are specifically challenging, as recapitulating hyaline cartilage can be difficult, thus compromising clinical outcomes. Progenitor cells for the treatment of articular cartilage injuries constitute a promising therapeutic method that has been increasing exponentially. Progenitor cells can be obtained from many different human tissues, such as bone marrow, adipose tissue, and muscle, as well as from peripheral blood after mobilizing stem cells from bone marrow with granulocyte colony-stimulating factor simulation. The minimally invasiveness, low complication rate, and efficacy of peripheral blood stem cells has gained significant attention and rapidly has become a promising source of progenitor cell delivery in the past decade.

See related article on page 2502

Cartilage injuries continue to be an important and common musculoskeletal problem in orthopaedic practices, where different treatment alternatives have been introduced in the past 50 years. The low intrinsic healing potential of cartilage tissue and the less-than-optimal fibrocartilage tissue obtained from several treatment options^{1,2} are the main limitations in achieving near-normal cartilage tissue after repair procedures.

Despite the promising results³⁻⁶ of current cartilage treatments, the ideal intervention is still a source of debate. This speaks to the lack of superiority among cartilage restoration treatments. When using adult chondrocytes, there are some concerns about their cell proliferation and differentiation capacity, as is the case

with autologous chondrocyte implantation.⁷ These issues have pushed clinicians and basic scientists to search for different treatment alternatives and cell sources that might be pluripotent in nature.

Although there are many issues that are still highly debated and over which there is no consensus, from the in vivo behavior to the way of its isolation and administration, and even to the mechanism of action and nomenclature, progenitor cells have already taken their place on the stage as the mysterious rock star of cartilage treatment because of their “potential” to differentiate into the native tissue. Following Arnold Kaplan’s statement,⁸ “they work, so use them,” with “why not?” clinicians have increasingly shown their motivation for the use of these cells in cartilage treatments in recent years. Furthermore, the use of cell-based therapies in the treatment of musculoskeletal pathologies has increased exponentially in the past decade.⁹

According to the pericyte theory,^{10,11} progenitor cells can be obtained from almost any human tissue. As such, progenitor cells are obtained from a variety of tissues, such as bone marrow, adipose, muscle, and peripheral blood.^{12,13} In this regard, progenitor cells obtained from peripheral blood are less invasive to obtain compared with bone marrow-derived cells and have a similar chondrogenic differentiation potential.¹⁴

Rush University Medical Center

The authors report the following potential conflicts of interest or sources of funding: J.C. reports other from Arthrex, ConMed Linvatec, Ossur, and Smith & Nephew, outside the submitted work; and board or committee member: American Orthopaedic Society for Sports Medicine; Arthroscopy Association of North America; and International Society of Arthroscopy, Knee Surgery, and Orthopaedic Sports Medicine: board or committee member. Full ICMJE author disclosure forms are available for this article online, as [supplementary material](#).

© 2021 by the Arthroscopy Association of North America
0749-8063/21375/\$36.00

<https://doi.org/10.1016/j.arthro.2021.03.026>

Peripheral blood stem cells (PBSCs) are based on the concept of mobilizing stem cells from the bone marrow with the administration of granulocyte colony-stimulating factor and collecting them from peripheral blood that is commonly used in other medical fields. PBSC technology has gradually increased since its first in vivo study¹⁵ in 2004, demonstrating promising results.¹⁶

We read the recent study by Saw, Anz, Ng, Jee, Low, Dorvault, and Johnson¹⁷ titled “Arthroscopic Subchondral Drilling Followed by Injection of Peripheral Blood Stem Cells and Hyaluronic Acid Showed Improved Outcome Compared to Hyaluronic Acid and Physiotherapy for Massive Knee Chondral Defects: A Randomized Controlled Trial” with great interest. We thank the authors for their elegantly executed study seeking to compare the effectiveness of PBSC plus hyaluronic acid after arthroscopic subchondral drilling with hyaluronic acid plus physiotherapy (control group) in a dual-center, randomized controlled trial for massive chondral defects of the knee joint. Randomized controlled trials in the biologic arena are scarce and, thus, we commend the authors in performing high-quality research in this field.

An important area in biologics is standardization in the reporting of cell-based treatments. To this point, the DOSES consensus outlined characteristics that needed to be reported to achieve maximal transparency as well as external validity of the studies involving progenitor cells.¹⁸ Saw et al.^{17,19-22} should be commended for continuing their line of research since 2005 regarding the effectiveness of PBSCs for the treatment of chondral injuries, as it has helped build on this novel concept consistently. In their previous studies, the authors reported that an 8-mL injection of fresh PBSCs on the seventh postoperative day contains an average of 20 million CD105+ cells. They also reported that the repair tissue obtained after PBSC treatment had hyaline cartilage histologic features.^{20,21} Second-look arthroscopic biopsy data containing detailed information about the microstructural features of the repair tissue within this large patient group would be beneficial.

A dose-dependent response of granulocyte colony-stimulating factor has been reported long before its use in orthopaedic practice, and its possible side effects are widely available in the literature.²³ Although the authors seem to have opted for a safe range by choosing a low dose, there are differences between the dose–response data reported previously²⁴ and the stem cell counts obtained in the current study. Randomized clinical trials with different cell dosage, with specific attention to the side effect profile, will further contribute to the delineation of the ideal treatment protocol. Unlike their previous study published in 2013,²¹ diagnostic arthroscopy and microfracture were not performed in the control group in this current study. Performing the same surgical

procedure on the control group would better demonstrate the isolated effect of PBSC independent of the microfracture procedure.

Of note, one important limitation to this procedure is ensuring patient compliance with the treatment, which includes 14 knee injections over the course of 18 months.²⁵ In addition, storage conditions, cost, and reduced cell viability in cryopreserved applications also can be listed as other concerns for the long treatment protocol.

It is also interesting that the study successfully visualized the microfracture holes and the cartilage tissue formed around them. The presented images in the recent study reveal the effect of the depth of microfracture holes and the distance between them on the treatment results and why we need novel microfracture methods. However, images of cartilage repair tissue formed only around the microfracture site can lead to believe that the scaffolding capacity of fibrin clots that forms in the microfracture defect area might not be ideal (as it has been previously reported²⁶ in lesions over 2 cm²). Thus, this might not create an optimal microenvironment in massive chondral injuries for a homogeneous distribution and proliferation of cells.

In conclusion, we congratulate the authors for their line of research on PBSCs and their recent well-designed randomized clinical study. International collaborations along with well-designed studies will allow us to advance the field. To this point, we encourage them to continue to further develop this concept as an alternative of cartilage repair.

References

1. Mow VC, Ratcliffe A, Rosenwasser MP, Buckwalter JA. Experimental studies on repair of large osteochondral defects at a high weight bearing area of the knee joint: A tissue engineering study. *J Biomech Eng* 1991;113:198-207.
2. Shapiro F, Koide S, Glimcher MJ. Cell origin and differentiation in the repair of full-thickness defects of articular cartilage. *J Bone Joint Surg Am* 1993;75:532-553.
3. Steadman JR, Briggs KK, Rodrigo JJ, Kocher MS, Gill TJ, Rodkey WG. Outcomes of microfracture for traumatic chondral defects of the knee: average 11-year follow-up. *Arthroscopy* 2003;19:477-484.
4. Hangody L, Dobos J, Balo E, Panics G, Hangody LR, Berkes I. Clinical experiences with autologous osteochondral mosaicplasty in an athletic population: A 17-year prospective multicenter study. *Am J Sports Med* 2010;38:1125-1133.
5. Andriolo L, Reale D, Di Martino A, et al. Long-term results of arthroscopic matrix-assisted autologous chondrocyte transplantation: A prospective follow-up at 15 years. *Am J Sports Med* 2020;48:2994-3001.
6. Chahla J, Sweet MC, Okoroha KR, et al. Osteochondral allograft transplantation in the patellofemoral joint: A systematic review. *Am J Sports Med* 2019;47:3009-3018.

7. Harrison PE, Ashton IK, Johnson WE, Turner SL, Richardson JB, Ashton BA. The in vitro growth of human chondrocytes. *Cell Tissue Bank* 2000;1:255-260.
8. Caplan AI. Medicinal signalling cells: They work, so use them. *Nature* 2019;566:39.
9. Rodeo SA. Cell therapy in orthopaedics: Where are we in 2019? *Bone Joint J* 2019;101-B:361-364.
10. Crisan M, Yap S, Casteilla L, et al. A perivascular origin for mesenchymal stem cells in multiple human organs. *Cell Stem Cell* 2008;3:301-313.
11. Caplan AI. All MSCs are pericytes? *Cell Stem Cell* 2008;3:229-230.
12. Han Y, Li X, Zhang Y, Han Y, Chang F, Ding J. Mesenchymal stem cells for regenerative medicine. *Cells* 2019;8.
13. Hernigou P, Homma Y, Flouzat-Lachaniette CH, Poignard A, Chevallier N, Rouard H. Cancer risk is not increased in patients treated for orthopaedic diseases with autologous bone marrow cell concentrate. *J Bone Joint Surg Am* 2013;95:2215-2221.
14. Wang SJ, Yin MH, Jiang D, et al. The chondrogenic potential of progenitor cells derived from peripheral blood: A systematic review. *Stem Cells Dev* 2016;25:1195-1207.
15. Jancewicz P, Dzienis W, Pietruczuk M, Skowronski J, Bielecki M. Osteochondral defects of the talus treated by mesenchymal stem cell implantation—early results. *Rocz Akad Med Białymst* 2004;49:25-27 (suppl 1).
16. Chen YR, Yan X, Yuan FZ, et al. The use of peripheral blood-derived stem cells for cartilage repair and regeneration in vivo: A review. *Front Pharmacol* 2020;11:404.
17. Saw KY, Anz AW, Ng RC, et al. Arthroscopic subchondral drilling followed by injection of peripheral blood stem cells and hyaluronic acid showed improved outcome compared to hyaluronic acid and physiotherapy for massive knee chondral defects: A randomized controlled trial. *Arthroscopy* 2021;37:2502-2517.
18. Murray IR, Chahla J, Safran MR, et al. International Expert Consensus on a Cell Therapy Communication Tool: DOSES. *J Bone Joint Surg Am* 2019;101:904-911.
19. Saw KY, Anz A, Jee CS, Ng RC, Mohtarrudin N, Ragavanaidu K. High tibial osteotomy in combination with chondrogenesis after stem cell therapy: A histologic report of 8 cases. *Arthroscopy* 2015;31:1909-1920.
20. Saw KY, Anz A, Merican S, et al. Articular cartilage regeneration with autologous peripheral blood progenitor cells and hyaluronic acid after arthroscopic subchondral drilling: A report of 5 cases with histology. *Arthroscopy* 2011;27:493-506.
21. Saw KY, Anz A, Jee CS, et al. Articular cartilage regeneration with autologous peripheral blood stem cells versus hyaluronic acid: A randomized controlled trial. *Arthroscopy* 2013;29:684-694.
22. Saw KY, Hussin P, Loke SC, et al. Articular cartilage regeneration with autologous marrow aspirate and hyaluronic acid: An experimental study in a goat model. *Arthroscopy* 2009;25:1391-1400.
23. Holig K. G-CSF in healthy allogeneic stem cell donors. *Transfus Med Hemother* 2013;40:225-235.
24. Kroger N, Zander AR. Dose and schedule effect of G-CSF for stem cell mobilization in healthy donors for allogeneic transplantation. *Leuk Lymphoma* 2002;43:1391-1394.
25. Posnett J, Dixit S, Oppenheimer B, Kili S, Mehin N. Patient preference and willingness to pay for knee osteoarthritis treatments. *Patient Prefer Adherence* 2015;9:733-744.
26. Bae DK, Song SJ, Yoon KH, Heo DB, Kim TJ. Survival analysis of microfracture in the osteoarthritic knee—minimum 10-year follow-up. *Arthroscopy* 2013;29:244-250.